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**Guide for authors**

Abstracts should be single spaced, aligned left with no justification, in Times New Roman 12 point font, with a 2 cm margin all around.

**Abstracts should be written in English and should not exceed 600 words** incl references Only extended abstracts by invited speakers: 5 pages inclusive references at maximum.

Do not use line breaks or format abstracts in columns.

Line 1: Title (bold and aligned left). Do not use abbreviations in the title.

Line 2: Name(s) of Author(s) (aligned left). Superscript 1, 2, etc. for address(es) different from that of first author. Name of presenting author should be underlined.

Line 3: Dept., Institution, City, State/Prov., Country, for each author, in sequence. (aligned left, no line breaks or line indents).

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Skip one line before body of text. No other blank lines are permitted. Text. (aligned left, no tabs, no indents, no line or column breaks).

Standard abbreviations may be used without definition. Other abbreviations must be defined by placing them in parentheses after the first time the full word appears. Basic methodology used and quantitative results must be included. Vague statements such as “additional results will be presented” should not be included.

References in the text (maximum of 3, extended abstracts by invited speakers: 10) should be indicated by superscript numbers in brackets corresponding with the reference list at the end of the abstract. Examples of references for a journal article and chapter in a book:

(1) Watts PD, Smith PA, Jouet LG, et al. Artificial insemination made easy. J Transatlan Vet Med 1985;14:28-35.

(2) Remlot BP and Luddy PW. Diseases of the severely immuno-deficient mouse. In: Weber RO, and Hedberg AC, eds. Fundamentals of transplantation surgery. Yorktown: WB Saunders Company, 1990:967-72.

Please stick to the file format of the following example:

**Prepubertal treatment with Suprelorin influenced uterine expression of GPR54 but not of KP-10, steroid hormone receptors and GnRH-R**

S. Schäfer-Somi1, D. Kaya2, M. Sözmen3, S. Kaya2, S. Aslan4

1Platform for Artificial Insemination and Embryo Transfer, Vetmeduni Vienna, Vienna, Austria 2Department of Obstetrics and Gynecology, Faculty of Veterinary Medicine, University of Kafkas, 36100, Kars, Turkey; 3Ondokuz Mayis University, Faculty of Veterinary Medicine, Department of Pathology, 55139 Atakum, Samsun  TURKEY 4Department of Obstetrics and Gynecology, Veterinary Faculty, Near East University,  Nicosia, North Cyprus, Turkey.

Email: Sab.lpdsc@gmail.com

Eleven crossbreed, pre-pubertal bitches aged 4.2±0.6 m were used as described in (1). Implants containing placebo (sodium chloride 0.9%; n = 4, G I), 4.7 mg (n = 3, GII) or 9.4 mg (n =4, GIII) deslorelin (Suprelorin; Virbac, France), were administered in the inter-scapular region (s.c). Signs of estrus and sexual behavioral changes were monitored once daily, until occurrence of estrus. Vaginal cytology, P4 and E2 concentrations were monitored every other day. Two bitches in GII and GIII came into estrus after 82.7±8.9 weeks after implantation and the controls after 61.9 ± 9.7 weeks. These animals were subjected to ovariohysterectomy 30-45 days later and uterine samples were collected. Some treated bitches (n=1 in G II and n=2 in G III) did not show estrus by the end of the observation period and were operated in week 101. Sections (5 μm) from the uterine tissue were subjected to immunohistochemistry (IHC) using the streptavidin-biotin peroxidase complex technique for detection of GnRH-R, Kisspeptin (KP)10, Kisspeptin receptor (GPR54), Androgen receptor (AR), Estrogen receptor (ER) alpha and beta, and Progesterone receptor (PR) as described by (2). Primary antibodies used for IHC: GPR54, 1/50, Novus Biologicals NLS1926; KP 10, 1/350, Merck Millipore, ab9754; GnRH-R, 1/15, Santa-Cruz, SC-8682; AR, 1/10, Santa-Cruz, SC-7305; ER alpha, 1/25, Abcam, ab32063; ER beta, 1/300, Biorbyt, orb83931; PR, 1/100, Biorbyt, orb11299. Tissue sections were scored semiquantitatively for immunolabelling. An immunoreactivity score (IRS) was calculated ranging from 0 to 300 (3). Results: No abnormalities were seen macroscopically or histologically in the uterine tissues. Kisspeptin 10 expression was low in all cell types. Highest IRS were seen in the vascular endothelial cells with highest IRS in G II (x=85). The GPR54 was mainly detected in the luminal epithelial cells, superficial and deep uterine glands, with maximum IRS in the luminal epithelial cells of G III (x=94.4). The expression increased with increasing Suprelorin concentration, especially when bitches were operated prepubertally (p<0.05). The PR, ER alpha and beta were exclusively expressed in superficial and deep uterine glands, and luminal surface epithelial cells but the distribution varied between individuals of all groups; in the latter, the ER alpha was higher in GII than in GI (p<0.05). The AR and GnRH-R expression was negative in all groups (GI-III) and all cell types, however positive in the control tissues (fetal trophoblast cells, midgestation canine uterus, ovarian luteal cells). We conclude that GPR54 expression was clearly influenced by prepubertal Suprelorin treatment but not PR, ER alpha and beta, AR, KP10 or GnRH-R.

(1) Kaya D, Schäfer-Somi S, Kurt B et al. Clinical use of deslorelin implants for the long-term contraception in prepubertal bitches: effects on epiphyseal closure, body development, and time to puberty. Theriogenology. 2015;83(7):1147-53.

(2) Schäfer-Somi S, Ay SS, Kaya D, et al.  Kisspeptin-10 and the G protein-coupled receptor 54 are differentially expressed in the canine pregnant uterus and trophoblast cells. Reprod Domest Anim. 2016;83(7):1147-53

(3) Sözmen M, Beytut E. An Investigation Of Growth Factors And Lactoferrin In Naturally Occurring Ovine Pulmonary Adenomatosis. J Comp Pathol. 2012;147(4):441-51